



WHO Policy Update: Interpretation of DST



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Technical Expert Consultation

FIND conducted systematic review of available minimum inhibitory concentration (MIC) data for **phenotypically wild type (pWT) strains**, as well as **genotypically non-wild type (gNWT) strains**, including strains from **allelic-exchange experiments** for second-line antituberculosis medicines.

The medicines included in the review were the **second-line injectable agents** (kanamycin, amikacin and capreomycin); clofazimine and bedaquiline; cycloserine and terizidone; linezolid; delamanid; and the **fluoroquinolones** (ofloxacin, levofloxacin, gatifloxacin and moxifloxacin).

Media reviewed: Löwenstein-Jensen, Middlebrook 7H10/7H11; BACTEC™ Mycobacterial Growth Indicator Tube™ 960 (MGIT).

Meeting Report

Technical Expert Group: critical concentrations for drug susceptibility testing for TB medicines

2017



Phenotypic methods for the diagnosis of DR-TB

Phenotypic, culture methods are based on assessment of the ability of *M. tuberculosis* to grow in culture media (solid or liquid) containing a critical concentration of specific anti-TB agents (which indicates **resistance**) or, conversely, its inability to grow in the same media (which indicates **susceptibility**).

The indirect proportion method is the most common method
Resistance is defined when at least 1% of growth is observed at the critical concentration of drug in the culture medium.

Commercial liquid culture systems for DST reduce the time to result to as little as 10 days, compared with the 28–42 days needed for DST using solid media

Revised definition: critical concentration

Critical concentration (CC) of an anti-tuberculous agent has been adopted and modified from international convention. The critical concentration is defined as the lowest concentration of an anti-TB agent that will inhibit the growth of at least 95% of wild-type strains of *M. tuberculosis*.

The critical concentration is typically the same or one dilution higher than the epidemiological cut-off value in order not to misclassify phenotypically wild-type strains as phenotypically non wild-type.

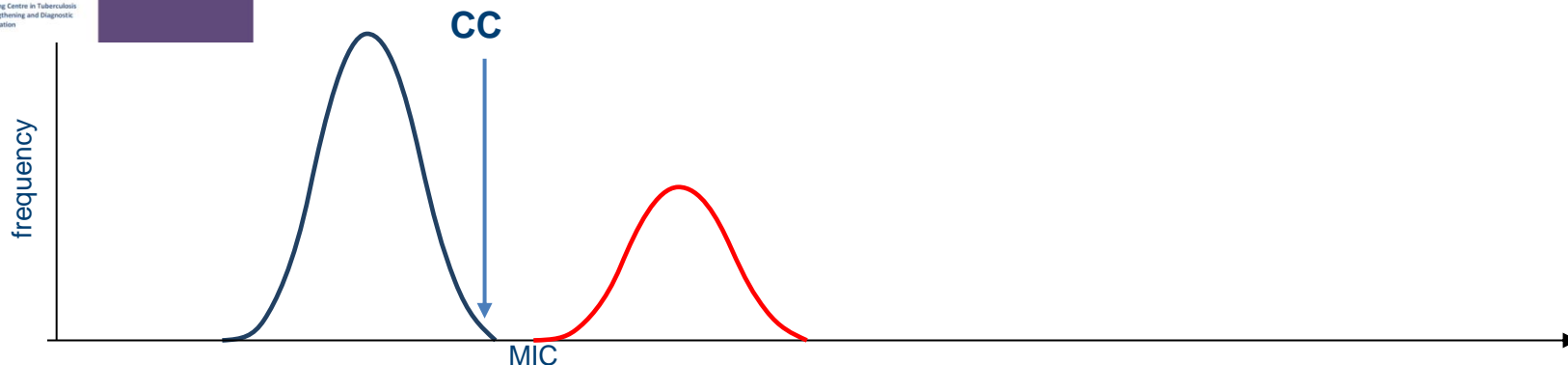
2017

Meeting Report

Technical Expert Group: critical concentrations for drug susceptibility testing for TB medicines



WHO Collaborating Centre in Tuberculosis Laboratory Strengthening and Diagnostic Technology Evaluation

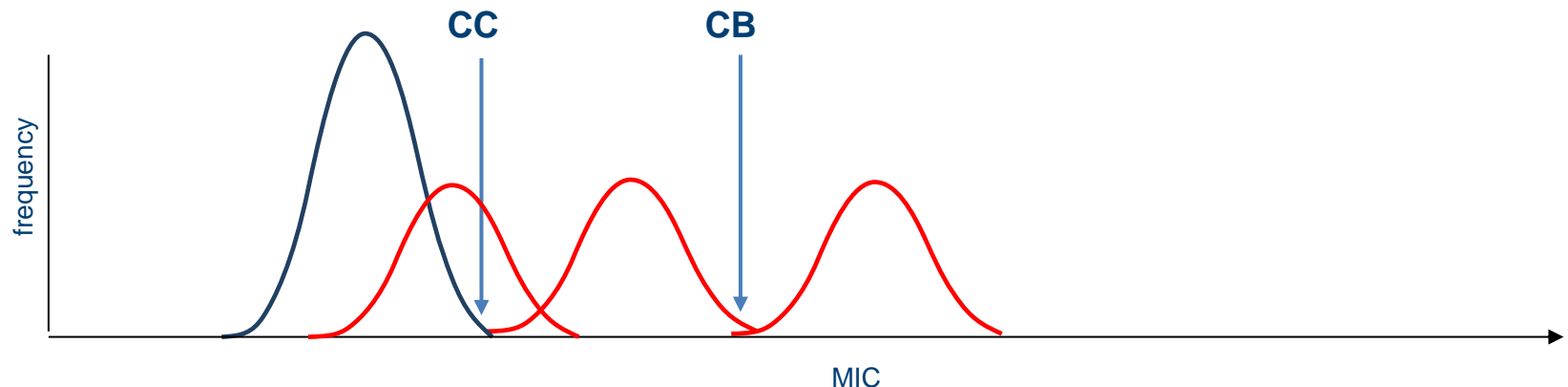


New definition: clinical breakpoint

Clinical breakpoint (CB)- is the concentration or concentrations of an antimicrobial agent which defines **an MIC which separates strains which will likely respond to treatment from those which will likely not respond to treatment**. This concentration is determined by correlation with available clinical outcome data, MIC distributions, genetic markers, and PK/PD data including drug dose.

A dose increase can be used to overcome resistance observed at lower dosing, up until the maximum tolerated dose, and therefore a higher clinical breakpoint above which the particular drug is not recommended for use.

The **clinical breakpoint** is commonly **different to the critical concentration** generally recommended for a given agent.



Phenotypic wild type (pWT) e.g. amikacin

MIC distributions on MGIT media

15 studies were identified that reported AMK MIC data for the pWT population with MGIT (Table 23). All studies had MICs for more than 10 pWT isolates. The MIC distributions reaffirmed the CC of 1 mg/L in MGIT.

Studies	Lab	Isolate origin	Unique isolates	Total MICs	Type of isolates	Sequencing results	AMK MIC [mg/L]																																
							0.06	0.12	0.25	0.5	1	1.5	2	2.5	4	8	16	20	32	64	128	256	512	1024															
21) Rüsçh-Gerdes 2006	15-17	clinical	10	30	H37Rv ATCC 27294 & pan-S								30																										
	15-17		21	63		different levels of R									36	9		3	15																				
	18		1	1			H37Rv ATCC 27294								1																								
22) Rodrigues 2008	18	clinical	10	10	pan-S								10																										
	18		20	20	different levels of R								16	2		1	1																						
	19		1	2	H37Rv																																		
23) Gonzalo 2015	19	clinical	20	40																																			
	19		20	40		gWT					1	20	19																										
24) Sturegård 2015	3	clinical	1	4	H37Rv ATCC 27294																																		
	3		28	28									8	12	3																						1	1	3
25) Heyckendorf	15	clinical	1	1	H37Rv ATCC 27294																																		
	15		9	9	MDR or XDR									2	7																								
26) Tessema 2017	15	clinical	40	40																																			
27) Zimencov 2013	20	clinical	1	1	H37Rv ATCC 25618																																		
	20		33	33										20	10	3																							
28) Kambli 2016a & 2016b	18	clinical	1	1	H37Rv ATCC 27294																																		
	18		31	31											25	5			1																				
29) Matt 2012	21	clinical	10	10	pan-S																																		
	22		1	3	H37Rv ATCC 27294																																		
30) Lin 2009	23	clinical	1	14																																			
	23		29	29																																			
31) Zheng 2016	11	clinical	1	1	H37Rv ATCC 25618																																		
	11		207	207	MDR																																		
32) Sharma 2011	24	clinical	36	36	different levels of R																																		
33) Cambau 2015	1, 3, 7, 15, 21, 24-27	clinical	113	113	MDR																																		
	1, 3, 7, 15, 21, 24-27		3	3																																			
34) Sirgel 2012	28	clinical	1	1	H37Rv ATCC 27294																																		
	28		15	15	MDR, XDR																																		
35) Springer 2009	21	clinical	11	11																																			

The green line denotes the current WHO and CLSI CC for AMK DST by MGIT (1 mg/L). Notable limitations: Only studies 23, 30-32 and 34 have data from unique laboratories.

Systematic review performed by Claudio Köser

Phenotypic wild type (pWT) moxifloxacin MIC distributions on MGIT media

Studies	Lab	Isolate origin	Unique isolates	Total MICs	Type of isolates	Genotypic results	0.06	0.12	0.18	0.25	0.5	0.75	1	1.5	2	2.5	3	4	5	7.5	8
23) Isaeva 2013, Nosova 2013 & Zimenkov 2013	17		1	1	H37Rv ATCC 25618			1													
	17	clinical	11	11	different levels of R	gWT	1	9		1											
	17		23	23			1	18		4											
24) Piersimoni 2007	18		20	20	H37Rv ATCC 27294 & pan-S		5	11		3	1										
	18	clinical	10	10			2	5		2			1								
	18		1	1		gWT									1						
25) Kambli 2015	19		1	1	H37Rv ATCC 27294		1														
	19	clinical	30	30		gWT	26	2		2											
26) Heyckendorf	20		1	1	H37Rv ATCC 27294					1											
	20	clinical	16	16	MDR or XDR	gWT	9	7													
27) Tessema 2017	20		41	41		gWT	8	32		1											
28) Rigouts	2		1	5	H37Rv ATCC 27294			4	1												
	2	clinical	9	9		gWT		9													
29) Sirgel 2012	9		1	6	H37Rv ATCC 27294			6													
	9	clinical	125	125	different levels of R	gWT		119		5						1					
30) van Ingen 2010	8		1	1	H37Rv ATCC 27294			1													
	8	clinical	28	28	MDR			20		1			1		6						
31) Krüüner 2006	21		132	132	MDR			97			5		16		14						
34) Ismail	6		57	57	different levels of R			26		12	2		2		12				2		1
	6	clinical	3	3		gWT		2					1								
33) Lin	22		73	73		gWT		44		22	1	1	1	3	1						
	22	clinical	218	218		gWT				215			2		1						
34) Cambau 2015	3, 8, 20, 23-28		114	114	MDR					114											
	3, 8, 20, 23-28	clinical	3	3		gWT				3											
35) Sharma 2011	29-32		36	144	different levels of R					101	6		12		25						
36) Kam 2006	33		108	108	mostly MDR						108										
	33	clinical	4	4		gWT							2		2						
37) Rodrigues 2008	19		1	1	H37Rv ATCC 27294								1								
	19		10	10	pan-S								10								
	19	clinical	20	20	different levels of R								19		1						
38) Alvarez 2014	34		1	1	H37Rv					1											
	34	clinical	5	5	MDR or XDR	gWT				1	1		2		1						

The blue line denotes the current CLSI CC for MFX DST by MGIT (0.25 mg/L). The red lines denote the current WHO CCs for MFX DST by MGIT (0.5 and 2 mg/L).

Systematic review performed by Claudio Köser



Table 1. Medicines recommended for the treatment of rifampicin-resistant and multidrug-resistant TB

Drug groups			LI proportion	10	7H11	MGIT 960	
A. Fluoroquinolones ⁽²⁾	Levofloxacin (CC)	Lfx		1.0	-	1.0	
	Moxifloxacin (CC) ⁽³⁾	Mfx	1.0	0.5	0.5	0.25	
	Moxifloxacin (CB) ⁽³⁾			2.0	-	1.0	
	Gatifloxacin ⁽⁴⁾	Gfx	0.5	-	-	0.25	
B. Second-line injectable agents	Amikacin	Am	30	2.0	-	1.0	
	Capreomycin	Cm	40	4.0	-	2.5	
	Kanamycin	Km	30	4.0	-	2.5	
	(Streptomycin)	(St)	4.0	2.0	2.0	1.0	
C. Other second-line agents	Ethionamide	Eto	40	5	10	5	
	Prothionamide	Pto	40	-	-	2.5	
	Cycloserine / Terizidone ⁽⁵⁾	Cs / Trd	-	-	-	-	
	Linezolid	Lzd	-	1.0	1.0	1.0	
	Clofazimine	Cfz	-	-	-	0.5	
D. Add-on agents (not part of the core MDR-TB regimen)	D1	Pyrazinamide	Z	-	-	-	100
		Ethambutol	E	2.0	5.0	7.5	5.0
	D2	Bedaquiline ⁽⁶⁾	Bdq	-	-	0.25	1.0
		Delamanid ⁽⁷⁾	Dlm	-	-	0.016	0.06
	D3	p-aminosalicylic acid	PAS	-	-	-	-
		Imipenem-cilastatin	Ipm	-	-	-	-
		Meropenem	Mpm	-	-	-	-
		Amoxicillin-clavulanate	Amx-Clv	-	-	-	-
		(Thioacetazone)	(T)	-	-	-	-

CC Critical concentration; CB Clinical breakpoint

Molecular methods for the diagnosis of DR-TB

Molecular (genotypic) methods **detect specific DNA mutations** in the genome of the *M. tuberculosis*, which are associated with resistance to specific anti-TB drugs.

Molecular methods have considerable advantages for **programmatic management of drug-resistant TB**, in particular with regard to their **speed**, the **standardization of testing**, their **potentially high throughput** and the reduced requirements for **laboratory biosafety**.

Molecular tests for detecting drug resistance to **rifampicin** alone or in combination with **isoniazid** have been recommended for use by WHO since 2008

Molecular methods for the diagnosis of DR-TB - limitations

There remains imperfect correlation between phenotypic and genotypic methods.

Molecular methods had high specificity but lower sensitivity which varies for different drugs

Rifampicin – *rpoB* 95% sensitivity, 99% specificity

Isoniazid – *inhA* and *katG* ~90% sensitivity, 99% specificity

Fluoroquinolones – *gyrA* and *gyrB* ~86% sensitivity, 99% specificity





Secondline injectable agents – *rrs* and *eis* ~86% sensitivity, 99% specificity

The predictive values of imperfect tests depend on the pre-test probability of resistance

The ReSeqTB Solution: A Standardized System for Grading Mutations

Observed frequency of a mutation found in phenotypically resistant and susceptible strains

- Literature data were used to calculate the frequency of each mutation
- Likelihood ratio (LR) and odd ratio (OR) were used; p-values and 95% confidence intervals associated with LR and OR have been also considered
- Thresholds used most commonly in evidence-based medicine have been adapted to grade *M. tuberculosis* mutations:

LR+ v OR p- adj value	Interpretation	Symbol
<0.05 > 10	High Confidence for association with resistance – strong association of the mutation with phenotypic drug resistance; sufficient evidence that the mutation confers or is strongly associated with drug resistance	
<0.05 5 < ... ≤ 10	Moderate Confidence for association with resistance – moderate association of the mutation with phenotypic drug resistance; additional data desirable for improved evidence that the mutation confers or is strongly associated with drug resistance	
<0.05 1 < ... ≤ 5	Low Confidence for association with resistance – weak association of the mutation with phenotypic drug resistance; inconclusive evidence that the mutation confers or is strongly associated with drug resistance. Substantial additional data required	
<0.05 < 1	No association with resistance – No evidence of association between the mutation and drug resistance	
≥0.05 -	Indeterminate – no statistically significant threshold reached; additional data required	Indeter

Slide courtesy of Paolo Miotto

NWT MFX MIC distributions of gyrA clinical mutants by MGIT

Studies	Medium	Total MICs	Genotypic results	MFX MIC (mg/L)																
				0.12	0.18	0.25	0.5	0.75	1	1.5	2	2.5	3	4	5	7.5	8	10		
23) Isaeva 2013, 25) Kambli 2015	MGIT	6	gyrA D94A				2		3		1									
28) Rigouts		4	gyrA D94A						1			3								
29) Sirgel 2012		11	gyrA D94A			3	5		3											
34) Ismail		7	gyrA D94A						4			3								
33) Lin		1	gyrA D94A				1													
33) Lin		6	gyrA D94A				1	3	1	1										
33) Lin		3	gyrA D94A				1		2											
34) Cambau 2015		2	gyrA D94A										2							
36) Kam 2006		5	gyrA D94A				1		4											
33) Lin		1	gyrA D94F						1											
23) Isae		13	gyrA D94G										13							
24) Pier		2	gyrA D94G									2								
25) Karr		42	gyrA D94G							9								1		
28) Rigc		20	gyrA D94G				1		9		10									
29) Sirg		17	gyrA D94G						3			14								
34) Ismail		2	gyrA D94G									1				1				
33) Lin		15	gyrA D94G							2	5	4			2	2				
33) Lin		16	gyrA D94G							1		13			1	1				
34) Cambau 2015		7	gyrA D94G										6				1			
36) Kam 2006		12	gyrA D94G									11			1					
38) Alvarez 2014	4	gyrA D94G							2		2			1						
34) Cambau 2015	1	gyrA G88C, D94G										1								
34) Cambau 2015	1	gyrA A90V, D94G															1			
29) Sirgel 2012	2	gyrA A90V, D94G							1		1									
23) Isaeva 2013,	1	gyrA S91P, D94N									1									
23) Isaeva 2013,	1	gyrA D94H									1									
25) Kambli 2015	3	gyrA D94H							1			2								
28) Rigouts	1	gyrA D94H									1									
33) Lin	4	gyrA D94H									2	2								
34) Cambau 2015	1	gyrA D94H										1								
36) Kam 2006	1	gyrA D94H										1								
23) Isaeva 2013,	1	gyrA D94N												1						
28) Rigouts	1	gyrA D94N										1								
29) Sirgel 2012	6	gyrA D94N					1				4			1						
34) Ismail	2	gyrA D94N							1		1									
33) Lin	1	gyrA D94N												1						
33) Lin	6	gyrA D94N									2			2	2					
34) Cambau 2015	1	gyrA D94N										1								
29) Sirgel 2012	2	gyrA D94G, D94N										2								
23) Isaeva 2013,	1	gyrA D94Y																		
28) Rigouts	1	gyrA D94Y																		
34) Ismail	1	gyrA D94Y																		

D94G mutations

Systematic review performed by Claudio Köser

Addressing current knowledge gaps

Drug	Gene	Mutation	Confidence value	
			Liquid DST	Solid DST
CM	<i>rrs</i>	a1401g	●	●
H	<i>inhA</i>	c-15t	●	●
KM	<i>eis</i>	g-10a	●	●
R	<i>rpoB</i>	L533P	indeter	●
R	<i>rpoB</i>	D516Y	indeter	●
MFX	<i>gyrA</i>	A90V	●	●
R	<i>rpoB</i>	L511P	●	●

Cases testing differently on different media

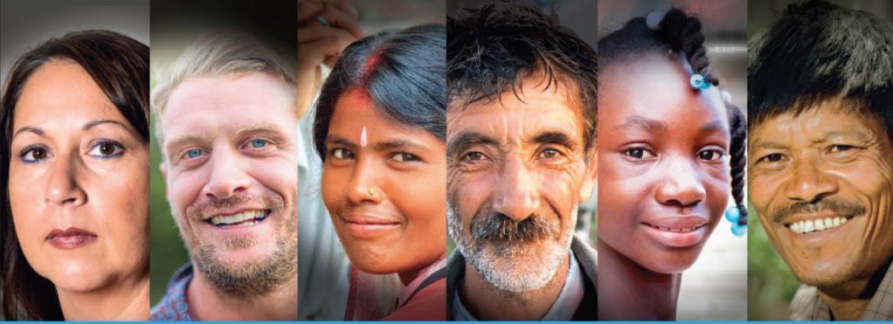
MIC close to liquid testing critical concentration

Cases missed by liquid phenotypic testing

Slide courtesy of Paolo Miotto

Addressing current knowledge gaps

1. Correlation of phenotypic DST critical concentrations with molecular methods – revised **critical concentrations are under review**
2. Incomplete cross resistance within the classes of key drugs such as the FQs – **mechanisms better understood**
3. **Technical guide** on the use of **DNA sequencing** as a **surveillance tool** and the **potential use and limitations** as a diagnostic tool – under development to determine mutations associated with elevated MICs for certain drugs
4. **PK/PD data** challenging our dogma around **suitable dosing** for key drugs **especially for moxifloxacin, rifampicin and pyrazinamide**



Compendium of WHO guidelines and associated standards:

ensuring optimum delivery of the cascade
of care for patients with tuberculosis

THE COMPENDIUM

END TB STRATEGY
POLICY
SCREENING
EARLIER DETECTION
DIAGNOSIS
TREATMENT
DRUG-RESISTANCE
MONITORING & EVALUATION
XDR-TB
TB/HIV
MDR-TB
TB IN CHILDREN
PALLIATIVE CARE
COMMUNITY ENGAGEMENT
LTBI
COMORBID CONDITIONS
PATIENT CARE AND SUPPORT
COMMUNITY
DIGITAL HEALTH
ETHICS
INFECTION CONTROL

THE
END TB
STRATEGY



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THANK YOU

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